

Local Coverage Determination (LCD): Foodborne Gastrointestinal Panels Identified by Multiplex Nucleic Acid Amplification Tests (NAATs) (L37350)

Links in PDF documents are not guaranteed to work. To follow a web link, please use the MCD Website.

Contractor Information

CONTRACTOR NAME	CONTRACT TYPE	CONTRACT NUMBER	JURISDICTION	STATE(S)
Noridian Healthcare Solutions, LLC	A and B MAC	01111 - MAC A	J - E	California - Entire State
Noridian Healthcare Solutions, LLC	A and B MAC	01112 - MAC B	J - E	California - Northern
Noridian Healthcare Solutions, LLC	A and B MAC	01182 - MAC B	J - E	California - Southern
Noridian Healthcare Solutions, LLC	A and B MAC	01211 - MAC A	J - E	American Samoa Guam Hawaii Northern Mariana Islands
Noridian Healthcare Solutions, LLC	A and B MAC	01212 - MAC B	J - E	American Samoa Guam Hawaii Northern Mariana Islands
Noridian Healthcare Solutions, LLC	A and B MAC	01311 - MAC A	J - E	Nevada
Noridian Healthcare Solutions, LLC	A and B MAC	01312 - MAC B	J - E	Nevada
Noridian Healthcare Solutions, LLC	A and B MAC	01911 - MAC A	J - E	American Samoa California - Entire State Guam Hawaii Nevada Northern Mariana

CONTRACTOR NAME	CONTRACT TYPE	CONTRACT NUMBER	JURISDICTION	STATE(S)
				Islands

LCD Information

Document Information

LCD ID

L37350

LCD Title

Foodborne Gastrointestinal Panels Identified by Multiplex Nucleic Acid Amplification Tests (NAATs)

Proposed LCD in Comment Period

N/A

Source Proposed LCD

DL37350

AMA CPT / ADA CDT / AHA NUBC Copyright Statement

CPT codes, descriptions and other data only are copyright 2018 American Medical Association. All Rights Reserved. Applicable FARS/HHSARS apply.

Current Dental Terminology © 2018 American Dental Association. All rights reserved.

Copyright © 2019, the American Hospital Association, Chicago, Illinois. Reproduced with permission. No portion of the AHA copyrighted materials contained within this publication may be copied without the express written consent of the AHA. AHA copyrighted materials including the UB-04 codes and descriptions may not be removed, copied, or utilized within any software, product, service, solution or derivative work without the written consent of the AHA. If an entity wishes to utilize any AHA materials, please contact the AHA at 312-893-6816. Making copies or utilizing the content of the UB-04 Manual, including the codes and/or descriptions, for internal purposes, resale and/or to be

Original Effective Date

For services performed on or after 02/11/2019

Revision Effective Date

For services performed on or after 10/01/2019

Revision Ending Date

N/A

Retirement Date

N/A

Notice Period Start Date

12/27/2018

Notice Period End Date

02/10/2019

used in any product or publication; creating any modified or derivative work of the UB-04 Manual and/or codes and descriptions; and/or making any commercial use of UB-04 Manual or any portion thereof, including the codes and/or descriptions, is only authorized with an express license from the American Hospital Association. To license the electronic data file of UB-04 Data Specifications, contact Tim Carlson at (312) 893-6816 or Laryssa Marshall at (312) 893-6814. You may also contact us at ub04@healthforum.com.

CMS National Coverage Policy

Title XVIII of the Social Security Act, §1862(a)(1)(A). Allows coverage and payment for only those services that are considered to be reasonable and necessary.

Title XVIII of the Social Security Act, §1833(e). Prohibits Medicare payment for any claim which lacks the necessary information to process the claim.

42 Code of Federal Regulations (CFR) 410.32(a). Diagnostic x-ray tests, diagnostic laboratory tests, and other diagnostic tests: Conditions.

CMS On-Line Manual, Publication 100-02, Medicare Benefit Policy Manual, Chapter 15, §§80.0, 80.1.1, 80.2. Clinical Laboratory services.

CMS Internet-Only Manuals, Publication 100-04, Medicare Claims Processing Manual, Chapter 16, §50.5 Jurisdiction of Laboratory Claims, 60.12 Independent Laboratory Specimen Drawing, 60.2. Travel Allowance.

CMS Internet Online Manual Pub. 100-04 (Medicare Claims Processing Manual), Chapter 23 (Section 10) "Reporting ICD Diagnosis and Procedure Codes".

Coverage Guidance

Coverage Indications, Limitations, and/or Medical Necessity

This contractor will provide limited coverage for Gastrointestinal Pathogen (GIP) molecular assays identified by multiplex nucleic acid amplification tests (NAATs), and will limit GIP coverage in immune competent beneficiaries up to 5 bacterial targets which represent the top 90-95% of foodborne infections ([incidence of infection per 100,000 population]in decreasing incidence): Salmonella [15.89]; Campylobacter [12.97]; Shigella [5.53]; Cryptosporidium [3.31]; Shiga toxin producing E. coli (STEC) non-O157 [1.64] and STEC O157 [.95].

In addition, when there is a clinical concern for Clostridium difficile colitis, this contractor will cover up to 11 targets if Clostridium difficile is one of the organisms tested for.

Testing for 12 or more organisms will only be covered in critically ill or immunosuppressed patients.

In immune competent individuals, most people with Cryptosporidium, a parasitic disease, will recover without treatment. The pathogens in some of the GIP panels are determined by the manufacturers that make them, and do not represent specific pathogens that cause a common age-based syndrome, or represent organisms that commonly

are found in a specific sample type, patient population or reflect community acquired foodborne infections. Because of the unique clinical circumstances of immune compromised patients, ICU patients, and HIV positive patients with diarrhea, GIP testing for bacteria, virus and parasite testing may be indicated, and thus a Medicare benefit.

Summary of Evidence

Traditionally, stool testing algorithms required physicians to consider which specific pathogens that might be associated with individual cases of gastroenteritis, and choose a testing scheme that ensured that all the appropriate pathogens were targeted. In the setting of community-acquired diarrheal illness, large foodborne GIP testing panels for parasites and viral etiologies is not reasonable and necessary because these GI diseases are:

- Generally self-limited,
- Virus specific therapies are not available, and
- Patients are managed by supportive care and hydration.

Travelers with >2 weeks of symptoms, after bacterial pathogens have been ruled out, may require traditional ova and parasite stool examination and/or specific protozoa antigen or molecular testing. Medicare specifies that testing must be reasonable and necessary for the specific needs of a given patient. Large panels that represent a “one size fits all” approach to testing without regard for a patient’s medical history, time of year, clinical setting, and patient symptoms are not reasonable and necessary, and thus not a Medicare benefit. A “one size fits all” panel approach is not restricted to specific population subgroups, such as neonates, pediatrics, or adults, does not differentiate between community-acquired vs traveler source of infection, and does not differentiate the needs of select patient populations such as the ICU patient or immunocompromised patients. In addition, while identification of specific viruses may be of interest in an outbreak or epidemiologically, clinical management is not predicated on viral test results, and are thus not reasonable and necessary.

This contractor recognizes that GIP assays are closed systems, without random access for physician directed, patient-specific testing. However, some laboratories elect to use GIP panel tests but report only the specific tests ordered by the physician. In other words, and the laboratory “blinds” unnecessary test results or utilize disclaimers in their reporting and bill only for the medically necessary test results. Other laboratories report results of all tests in the panel which adds unnecessary cost to the healthcare system when reimbursement is directly related to the number of organisms in the panel. The FDA approved/cleared assays discussed below are comparable with coverage limited to bacterial organisms for acute diarrhea, with justification of medical necessity recorded in the patient’s medical record.

Nucleic Acid Amplified Probe Technique (NAAT) for Identification of Microorganisms:

Tests performed by NAAT uses a microorganism’s DNA or RNA to directly identify specific bacteria, viruses, and/or protozoa rather than standard microorganism detection techniques such as bacterial culture, microscopy with and without stains, direct fluorescent antibody testing, rapid antigen testing, qualitative and quantitative immunoassay for identification of antigens or toxins from stool and single-plex PCR assays. Multiplex NAAT tests are included in the larger grouping of culture-independent diagnostic tests (CIDT). CIDT includes but is not limited to simplex direct probe and amplified probe techniques. This technology offers same day results in a matter of hours rather than 2-3

days of time-consuming and labor intensive bacterial cultures and immunoassays for processing stool specimens. CIDT are touted as providing a more comprehensive assessment of disease etiology by increasing the diagnostic yield compared with conventional diagnostic tests permitting earlier initiation of appropriate therapeutic agents targeted to the detected pathogen(s), if any, rather than empirical therapy until culture results are available.

CIDT testing is not without its challenges; latent infections or colonization cannot be distinguished from active, clinically significant infections. Additionally, fragments of nucleic acids from dead microorganisms may cloud organism identification, complicating clinical interpretation, and potentially, clinical management. In a CIDT comparative study, mixed infections were identified in 13-21% of positive prospective stool samples compared to only 8.3% by routine (culture/immunoassay/microscopy) methods.¹ In another recent study, 32.9% of the FilmArray GI Panel-positive specimens were found to contain more than one potential pathogen.² The significance of detecting coinfections may be difficult to understand, as the clinical implications of specific pathogen combinations are not well documented or understood. Many GI pathogens can be shed asymptotically or for prolonged periods of time after symptoms subside, further complicating the interpretation of positive results. For example, *Salmonella* spp. and norovirus can be shed for weeks to months after symptoms subside. Asymptomatic infection with *Cryptosporidium* spp. or *G. lamblia* is common in children.² High rates of asymptomatic carriage of enteropathogens, often identified as a co-infection in large microbial panels, create diagnostic confusion by the interpreting clinician.³

From a public health and epidemiologic point of view, CIDT testing does not provide the culture isolates that are needed for antimicrobial susceptibility testing, serotyping, subtyping and whole genome sequencing that are critical for monitoring trends, detecting clusters of illness and investigating outbreaks. For *Salmonella*, the inability to distinguish serotypes will prevent tracking of important changes in incidence by serotype, and markedly limit detection and investigation of outbreaks (not a Medicare benefit). For Shiga toxin producing *E. coli* (STEC), because identification of serogroups requires culture, it is not known which STEC-positive CIDT result represents 0157 vs non-0157⁴.

FDA-approved GIP Assays:

Five FDA approved GIP assays are currently on the market, and all are closed system tests that do not allow random access for physicians to select likely etiologic agents of diarrhea. These include:

- **Hologic/Gen-Probe's ProGastro SSCS**

- **Targets identified:**
 - *Salmonella*,
 - *Shigella*,
 - *Campylobacter* (*C. jejuni* and *C. coli* only, undifferentiated) nucleic acids, and
 - Shiga toxin 1 (stx1) /Shiga toxin 2 (stx2) genes (STEC typically harbor one or both genes that encode for Shiga toxins 1 and 2)
- TAT (turn-around time) - 4 hr.

- **BD Diagnostics' BD MAX Enteric Bacterial Panel (EBP):**

- Targets identified:
 - *Campylobacter* spp. (*jejuni* and *coli*),
 - *Salmonella* spp.,
 - *Shigella* spp.,
 - Enterohemorrhagic *E. coli* (EHEC),
 - Shiga toxin 1 (stx1)/Shiga toxin 2 (stx2) genes (found in STEC, as well, as *Shigella dysenteriae*)
- TAT – 3-4 hr.

- **Nanosphere's Verigene Enteric Pathogens (EP):**

- Targets identified:
 - *Campylobacter* Group (comprised of *C. coli*, *C. jejuni*, and *C. lari*),

- Salmonella species,
 - Shigella species (including *S. dysenteriae*, *S. boydii*, *S. sonnei* and *S. flexneri*),
 - Vibrio Group (comprised of *V. cholera* and *V. parahaemolyticus*),
 - Yersinia enterocolitica,
 - Shiga toxin I gene and Shiga toxin 2 gene virulence markers, Shiga toxin producing E coli (STEC)
 - Norovirus
 - Rotavirus
- TAT – 2 hr.
- **Luminex’s xTAG Gastroenterology Pathogen Panel (GPP):**
 - Targets identified
 - Campylobacter (*C. jejuni*, *C. coli* and *C. lari* only)
 - Clostridium difficile (*C. difficile*) toxin A/B
 - Cryptosporidium (*C. parvum* and *C. hominis* only)
 - Escherichia coli (*E. coli*) O157
 - Enterotoxigenic E. coli (ETEC) LT/ST
 - Giardia (*G. lamblia* only) (aka *G. intestinalis* and *G. duodenalis*)
 - Norovirus GI/GII
 - Rotavirus A
 - Salmonella
 - Shiga-like Toxin producing E. coli (STEC) stx 1/stx 2
 - Shigella (*S. boydii*, *S. sonnei*, *S. flexneri* and *S. dysenteriae*)
 - *E. histolytica*
 - Adenovirus 40/41
 - Vibrio cholera
 - TAT - <5 hr.
- **Biofire Diagnostic’s FilmArray GI Panel:**
 - Targets identified
 - Campylobacter (*C. jejuni*/*C. coli*/*C. upsaliensis*),
 - Clostridium difficile (*C. difficile*) toxin A/B,
 - Plesiomonas shigelloides,
 - Salmonella,
 - Vibrio (*V. parahaemolyticus*/*V. vulnificus*/ *V. cholerae*), including specific identification of *Vibrio cholerae*,
 - Yersinia enterocolitica,
 - Enteroaggregative Escherichia coli (EAEC),
 - Enteropathogenic Escherichia coli (EPEC),
 - Enterotoxigenic Escherichia coli (ETEC) lt/st,
 - Shiga-like toxin-producing Escherichia coli (STEC) stx1/stx2 (including specific identification of the *E. coli* O157 serogroup within STEC),
 - Shigella/ Enteroinvasive Escherichia coli (EIEC),
 - Cryptosporidium,
 - Cyclospora cayetanensis,
 - Entamoeba histolytica,
 - Giardia lamblia (also known as *G. intestinalis* and *G. duodenalis*),
 - Adenovirus F 40/41,
 - Astrovirus,
 - Norovirus GI/GII,
 - Rotavirus A,
 - Sapovirus (Genogroups I, II, IV, and V)
 - TAT -1-2 hr.

All targeted viruses included in GIPs are more prevalent in young children than in adults. In one study, Sapovirus was detected in 10% of all specimens from children >1 year old and 7.4% of specimens from children between 1 to 5 years of age.²

Enteropathogenic *E. coli* (EPEC), historically associated with developing countries, are known to cause both acute and persistent diarrhea in 2 young children in the US and were identified in one study in 24.8% of all samples collected from children <1 year of age, and 37% of all samples from children between age of 1 and 5 years. EPEC strains can also be found in healthy children and adults, thus confounding its significance when identified in symptomatic children and adults.

Similarly, the interpretation of *C. difficile* toxin A/B detection is also complicated, especially in children <1 year old. The American Academy of Pediatrics does not recommend routine testing for *C. difficile* in children <1 year of age and suggests that positive *C. difficile* results be interpreted with suspicion in children <3 years old⁵.

Most recently a publication looking at Syndromic Panel-Based Testing, specifically the multiplex detection of GI pathogens, states that it makes it difficult the interpretation of a positive results in asymptomatic individuals colonized with *C. difficile*. Patients experiencing diarrhea associated with antecedent of antibiotic or hospitalization are at risk for *C. difficile* infection; in such cases specific testing for *C. difficile* is most cost-effective. GIP's detection of multiple targets has created confusion for healthcare providers that now faced with results that were not previously reported and for which current guidelines provide no direction as to management (treatment, clinical significance or the need for additional or repeat testing)¹².

A meta-analysis (of 10 studies) by NHS in UK found that GIP testing produces a greater number of pathogen-positive findings than conventional testing. It is unclear whether or not these additional "positives" are clinically important. The review identified no robust evidence to inform consequent clinical management of patients. There is considerable uncertainty about cost-effectiveness of GIP panels used to test for suspected infectious gastroenteritis in hospital and community settings. The systemic review and cost-effectiveness model identify uncertainties about the adoption of GIP tests. GIP testing will generally identify pathogens identified by conventional testing, however, these tests also generate considerable additional positive results of uncertain clinical importance¹³.

Indications for Foodborne GI Testing

Acute diarrhea, often called gastroenteritis, can be defined as the passage of a greater number of stools of decreased form from the normal lasting < 14 days. Acute diarrhea is generally associated with clinical features of nausea, vomiting, abdominal pain and cramps, bloating, flatulence, fever, passage of bloody stools, tenesmus and fecal urgency. It is the leading cause of outpatient visits, hospitalizations, and lost quality of life occurring domestically and those traveling abroad. The Centers for Disease Control and Prevention (CDC) has estimated 47.8M cases occurring annually in the US with an estimated healthcare cost upwards of US\$150M.⁶ Detection of microbial pathogens associated with GI disease may be important in certain populations, such as immunocompromised hosts, the critically ill and individuals with prolonged disease that is refractory to treatment.

Over a 20 year period, some foods that have been linked to food-borne outbreaks including milk (*Campylobacter*), shellfish (Noroviruses), unpasteurized apple cider (*Escherichia coli* O157:H7), raw and undercooked eggs (*Salmonella*), fish (ciguatera poisoning), raspberries (*Cyclospora*); strawberries (*Hepatitis A virus*); and ready-to-eat meats (*Listeria*).⁷

Although the etiologic agents responsible for about 80% of GI illnesses are unidentified or otherwise unspecified, Norovirus and *Salmonella* spp (non-typhoidal) are currently the most commonly identified pathogens associated with

food-borne disease in the US and account for 5.5 and 1.0 million cases each year, respectively.⁸ Clostridium perfringens, Campylobacter and Staphylococcus aureus follow Norovirus and Salmonella spp. in decreasing frequency in domestically acquired foodborne illnesses. Healthcare- and antibiotic-associated diarrhea is also problematic, with the major causative pathogen being toxin-producing Clostridium difficile⁹. In the US, >300,000 cases of C. difficile are diagnosed annually, with associated costs of >\$1 billion.

In 2015, the number and incidence of confirmed infections per 100,000 population were reported for Salmonella (15.89), Campylobacter (12.97), Shigella (5.53), Cryptosporidium (3.31), Shiga-toxin producing Escherichia coli (STEC) non-O157 (1.64), STEC O157 (.95), Vibrio (0.39), Yersinia (0.29), Listeria (0.24) and Cyclospora (0.13).⁴ Among confirmed infections, the vast majority were diagnosed only by culture. Compared with incidence in 2012-2014, the incidence of confirmed infections was significantly higher for STEC non-O157 (40% increase) and Cryptosporidium (57% increase). No significant changes were observed in 2015 for other pathogens compared with the previous 3-year averages. In addition to the 20,107 confirmed cases of infection, there were 3,112 positive CIDT case reports. In general, the incidence of most foodborne bacterial pathogens and for Cryptosporidium is highest among children aged <5, except for Listeria and Vibrio for which the highest incidence is among persons aged = 65 years.¹⁰

Many episodes of acute diarrhea are self-limited and require fluid replacement and supportive care. Oral rehydration is indicated for patients who are mildly to moderately dehydrated. IV fluids may be required for more severe dehydration. Routine use of antidiarrheal agents is not recommended because many of these agents have potentially serious adverse effects, particularly in infants and young children. Antimicrobial therapy is warranted only for patients with severe disease or for individuals with immune systems are severely weakened from medications and other illnesses.¹¹

Laboratory testing algorithms for infectious causes of diarrhea generally agree that testing is NOT warranted for community-acquired diarrhea of <7 days duration without signs or symptoms of severe (fever, bloody diarrhea, dysentery, severe abdominal pain, dehydration, hospitalization and immunocompromised state) disease. In general, when community-acquired diarrhea persists for =7 days, or the diarrhea is travel-related, or there are signs/symptoms of severe disease, GIP testing may be warranted. Additional directed testing may be indicated if the GIP results are negative and diarrhea persists. No additional testing is indicated for GIP-positive result unless the clinical pictures changes. Clostridium difficile molecular testing is warranted on health-care associated diarrhea with onset after the 3rd inpatient day or after recent antibiotic use.

Analysis of Evidence (Rationale for Determination)

Level of Evidence

Quality of Evidence: Moderate

Strength of Evidence: Moderate

Weight of Evidence: Moderate

Summary Medicare Coverage Decision:

GIP testing is limited to no more than 5 bacterial pathogen targets when not testing for *Clostridium difficile*. Testing for 6-11 pathogens is covered when there is a clinical concern for *Clostridium Difficile* colitis, and *Clostridium difficile* is one of the pathogens being tested.

Testing for viral etiologies is not reasonable and necessary because these GI diseases are generally self-limited, virus specific therapies are not available, and patients are managed by supportive care and hydration. Travelers with >2weeks of symptoms, after bacterial pathogens have been ruled out, may require traditional ova and parasite stool examination and/or specific protozoa antigen or molecular testing. Large panels inclusive of 11 viruses and protozoa are not reasonable and necessary for community-acquired diarrheal illness. There is no Medicare benefit for GIP testing for national, state or local agency tracking of diarrheal outbreaks, for epidemiologic purposes, or to confirm another etiologic test result. Once the target etiology of an outbreak is identified, subsequent patient testing is generally not indicated and patients are managed empirically. However, if the clinical presentation varies from the outbreak prototype, a specific test for the causative organism may be indicated. The Medicare benefit is specifically for the clinical identification and management of disease for a given beneficiary. The Medicare benefit does not extend for purposes of the family or community tracking or surveillance.

Limitations

A GIP test **panel** is a single service with a single unit of service (UOS =1). A panel cannot be unbundled and billed as individual components regardless of the fact that the GIP test reports multiple individual pathogens and/or targets. The panel is a closed system performed on a single platform, and as such, is a single test panel with multiple components (UOS=1). If *C. difficile* is not included in a GIP panel, testing for *C. difficile* may be reasonable and necessary when ordered in addition to a GIP bacterial pathogen panel and supported by documentation in the medical record.

General Information

Associated Information

N/A

Sources of Information

N/A

Bibliography

References

1. Khare R, Espy MJ, Celelinski E, et al. Comparative evaluation of two commercial multiplex panels for detection of gastrointestinal pathogens by use of clinical stool specimens. *J Clin Microbiol* 2014; 52(10):3667-73.
2. Buss SN, Leber A, Chapman K, et al. Multicenter evaluation of the BioFire FilmArray gastrointestinal panel for etiologic diagnosis of infectious gastroenteritis. *J Clin Microbiol*. 2015; 53:915-25 doi: 10.1128/JCM.02674-14.
3. Binnicker, MJ. Multiplex molecular panels for diagnosis of gastrointestinal infection: performance, result

- interpretation, and cost-effectiveness. J Clin Microbiol 2015; 53: 3723-8. doi:10.1128/JCM.02103-15.
4. Huang JY, Henao OL, Griffin PM, et al. Infection with pathogens transmitted commonly through food and the effect of increasing use of culture-independent diagnostic tests on surveillance – foodborne diseases active surveillance network, 10 US sites, 2012-2015. MMWR 2016; 65(14):369-71.
 5. Schutze GE, Willoughby RE, Committee on Infectious Diseases, American Academy of Pediatrics. Clostridium difficile infection in infants and children. Pediatrics 2013;131:196-200. <http://dx.doi.org/10.1542/peds>.
 6. Riddle MS, DuPont HL Connor BA. ACG Clinical Guideline: Diagnosis, treatment and prevention of acute diarrheal infections in adults. Am J Gastroenterol 2016; 111:602-22. doi:10.1038/ajg.2016.126.
 7. Center for Disease Control and Prevention. Diagnosis and management of foodborne illnesses: A primer for physicians and other health care professionals. MMWR 2004; 53,RR-4:1-33.
 8. Scallan E, Griffin PM, Angulo FJ, et al. Foodborne illness acquired in the United States– unspecified agents. Emerg Infect Dis 2011; 17:16–22. <http://dx.doi.org/10.3201/eid1701.P21101>
 9. Kelly CP and Lamont JT. Patient education: Antibiotic-associated diarrhea caused by Clostridium difficile (Beyond the Basics) UpToDate 2017; 25.2-C25.93.
 10. Incidence and trends of infection with pathogens transmitted commonly through food – foodborne diseases active surveillance network, 10 US sites, 1996-2012. MMWR 2013; 62(15):283-7.
 11. CDC, 2011 <https://www.cdc.gov/foodborneburden/2011-foodborne-estimates.html>.
 12. Ramanan P, et al. Syndromic panel-based testing in clinical microbiology. Clin Micro Rev 2018; January vol 31;issue 1;e00024-17.
 13. Freeman K, et al. Multiplex tests to identify gastrointestinal bacteria, viruses and parasites in people with suspected infectious gastroenteritis: a systemic review and economic analysis. Health Technol Assess 2017;21(23).

Revision History Information

REVISION HISTORY DATE	REVISION HISTORY NUMBER	REVISION HISTORY EXPLANATION	REASON(S) FOR CHANGE
10/01/2019	R4	10/01/2019: All coding located in the Coding Information section has been moved into the related Billing and Coding: Foodborne Gastrointestinal Panels Identified by Multiplex Nucleic Acid Amplification (NAATs) A56706 article and removed from the LCD.	<ul style="list-style-type: none"> • Revisions Due To Code Removal
02/11/2019	R3	08.19.19: Removal of codes per requirement and billing and coding article created to complement LCD and removal of MoIDX from title of LCD	<ul style="list-style-type: none"> • Revisions Due To ICD-10-CM Code Changes • Revisions Due To CPT/HCPCS Code Changes
02/11/2019	R2	12/21/2018: Removal of verbiage 'in addition to a diagnosis code from Group 1' from Group 2:Paragraph per Palmetto GBA.	<ul style="list-style-type: none"> • Creation of Uniform LCDs With Other MAC Jurisdiction

REVISION HISTORY DATE	REVISION HISTORY NUMBER	REVISION HISTORY EXPLANATION	REASON(S) FOR CHANGE
02/11/2019	R1	LCD revised to add E. histolytica, Adenovirus 40/41 and Vibrio cholera under FDA approved GIP Assays for Luminex's xTAG Gastroenterology Pathogen Panel (GPP) Target identified.	<ul style="list-style-type: none"> • Creation of Uniform LCDs With Other MAC Jurisdiction • Typographical Error

Associated Documents

Attachments

N/A

Related Local Coverage Documents

Article(s)

A56706 - Billing and Coding: Foodborne Gastrointestinal Panels Identified by Multiplex Nucleic Acid Amplification (NAATs)

A56205 - Response to Comments: MoIDX: Foodborne Gastrointestinal Panels Identified by Multiplex Nucleic Acid Amplification (NAATs)

LCD(s)

DL37350 - MoIDX: Foodborne Gastrointestinal Panels Identified by Multiplex Nucleic Acid Amplification Tests (NAATs)

Related National Coverage Documents

N/A

Public Version(s)

Updated on 09/20/2019 with effective dates 10/01/2019 - N/A

Updated on 08/19/2019 with effective dates 02/11/2019 - 09/30/2019

Updated on 12/26/2018 with effective dates 02/11/2019 - N/A

Updated on 12/20/2018 with effective dates 02/11/2019 - N/A

Updated on 12/11/2018 with effective dates 02/11/2019 - N/A

Keywords

- MoIDX
- Foodborne
- Gastrointestinal
- Multiplex
- Nucleic Acid
- Amplification
- NAATs